

Postnatal metabolic changes in the pups of rats exposed to toddy (palm wine) during pregnancy and lactation

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Female rats were exposed to toddy (palm wine) $(24.5 \text{ ml kg}^{-1} \text{ body weight day}^{-1})$ and ethanol $(0.52 \text{ ml kg}^{-1} \text{ body weight day}^{-1})$ before conception and throughout gestation and lactation. For 21 days pups were nursed by their own mothers, afterwards they were fed normal laboratory feed. Examination of the lipid profile of various tissues of 21 day-old pups revealed marked hyperlipidemia. The tissues of 45 day-old pups showed that even though there was some regression in the lipid accumulation, the status of hyperlipidemia prevailed. The lipogenesis was enhanced on both the days studied as evidenced by the increased activity of the lipogenic enzymes, HMG Co A reductase and incorporation of 14 C acetate to lipids. Catabolism of cholesterol to bile acids was decreased. The biochemical alterations produced in the pups by toddy and its equivalent in alcohol were different, showing that the non-alcoholic portion of the toddy interfered with the lipid metabolism. © 1998 Elsevier Science Ltd. All rights reserved

INTRODUCTION

Chronic ingestion of alcohol affects a variety of organ systems and most markedly the liver (Charls, 1980). Maternal exposure to alcohol leads to fetal alcohol syndrome or FAS (King and Fabro, 1983) which is characterised mainly by abnormalities of the nervous system (Clarren and Smith, 1978). Decreased density of synaptic formation was observed in the fetus and pups on exposure to ethanol during gestation (Tanaka et al., 1991). The degree of damage was dose-dependent (Norton and Kotkoskie, 1991) and it affected the development of the CNS (Norton et al., 1988). Most of the studies during the postnatal development of pups of alcohol-exposed dams were on behavioural problems (Driscoll et al., 1985; Abel and Dintcheff, 1986). Consumption of alcohol leads to hyperlipidemia and the mechanism of its formation was well documented (Jones, 1969). Very few studies were conducted to assess alterations in the lipid metabolism of the young of dams administered ethanol during gestation and lactation. Almost all the research work done in alcohol metabolism was carried out with pure ethyl alcohol. But in reality it is alcoholic beverages that are being consumed. Alcoholic beverages contain many pharmacologically active substances other than ethanol. Toddy, (palm wine) is a popular undistilled alcoholic beverage consumed mainly by the low socio-economic strata of the society in Kerala. It is procured from coconut trees (*Cocos nucifera*). Therefore it was decided to study lipid metabolism of pups of dams exposed to toddy/ethanol during gestation and lactation.

MATERIALS AND METHODS

Female albino rats (Sprague Dawley Strain) of weight 125g were randomnly selected and divided into three groups of 6 rats each as:

- 1. control (Laboratory diet);
- 2. toddy (24.5 ml kg $^{-1}$ body weight); and
- 3. ethanol $(0.52 \text{ ml kg}^{-1} \text{ body weight})$.

This dosage takes into consideration that a habitual drinker on average consumes 2 bottles (1250 ml) of toddy per day.

The rats were maintained in laboratory conditions in a light and dark schedule of 12 h duration. They were fed with pelleted diet (Lipton India Ltd) and tap water *ad libitum*. The group II animals were given 12 h-fermented toddy purchased daily from the same Government licenced shop and its alcohol content was estimated to be 2–2.5% (AOAC). Ethanol/toddy was administered by gastric intubation. The alcohol content

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of the ethanol group was adjusted to be equivalent to that in the toddy group. Ethanol was diluted with water in the ratio 1:7 and toddy was given without dilution. Control rats were fed with an equicaloric amount of glucose solution. After 15 days of treatment they were allowed to mate with normal males for four days. Pregnancy was detected by microscopical examination of Alcohol/toddy vaginal smear. was administered throughout gestation and lactation. The day of delivery was considered as the first day of lactation. Pups were nursed by their own mothers. On the 21st day of lactation, half of the pups were sacrificed by cervical dislocation. The rest of the animals were allowed to grow. They were fed with laboratory pelleted rat feed during this period. On 45th day they were sacrificed and various tissues and serum were collected into pre-cooled containers and analysed by standard procedures. Tissues were stored at $-4^{\circ}C$ for the analysis of lipids (Menon and Kurup, 1976) and bile acids (Okishio et al., 1967). Activities of enzymes were analysed immediately after tissue collection. Incorporation of ¹⁴C acetate to lipids was studied (Molly et al., 1983) as previously described. The activity of the enzymes glucose-6-phosphate dehydrogenase (EC 1.1.1.49) (Kornberg and Horecker, 1960), Malic enzyme (EC.1.1.1.40) (Severo, 1960) and HMG Co-A reductase (EC.1.1.1.34) (Rao and Ramakrishnan, 1975) were also assayed. Protein in the enzyme extract was determined by the method of Lowry et al. (1951). Statistical analysis and comparison between groups were done by Student's t test (Bennet and Franklin, 1967).

RESULTS

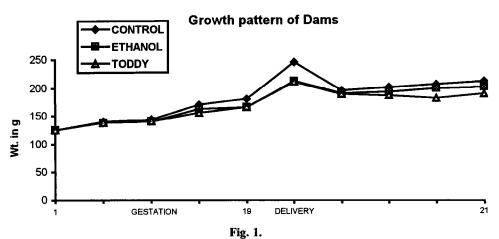
The dams treated with toddy and ethanol showed a significant decrease in the weekly weight gain, in comparison with the control group. The body weight on the day of delivery showed a significant reduction in the toddy treated group and it continued up to the last day of lactation (Fig. 1). Weights of the pups were also lower in the treated groups. A similar pattern in the weight gain was observed in the pups of all groups. On day 45, a drastic weight reduction was noted in the toddy group (Fig. 2).

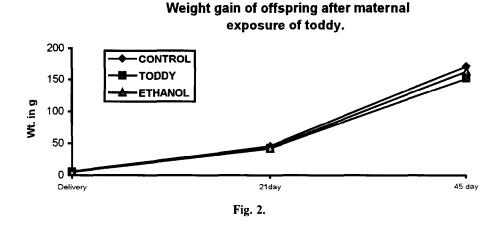
There was significant reduction in the number of pups born to the dams fed toddy/ethanol. There was a decrease in the survival of pups when they were fed by mothers milk, but it was not statistically significant. Afterwards there were no differences in the survival rates of the pups in the different groups (Fig. 3).

The cholesterol content in the liver and brain of 21st day pups showed a significant increase in both the treated groups. On the 45th day the cholesterol level came down to normal level in the liver of animals administered toddy. In the kidney the cholesterol content did not show any significant alterations in the treated groups in comparison to their control. In brain of 45th day pups, the total cholesterol showed a significant increase in toddy group, while there was no significant change in the ethanol-treated group (Table 1). There was no significant alteration in the total cholesterol content of the serum. However, HDL-C showed a significant reduction in the toddy/ethanol groups on the 21st and 45th days and LDL+VLDL-C showed a significant increase (Table 3).

The phospholipids showed an increase in the liver, brain and kidney of 21st day pups treated with toddy, but there was no change in animals exposed to ethanol. In 45th day pups, a significant increase in phospholipid was found only in the kidney of the treated groups. The serum phospholipids showed a marked increase in both the treated groups in comparison with controls. However, significant alterations were not observed between 21st and 45th day pups (Table 1).

The triglycerides showed a significant increase in all the pups administered toddy, but in the ethanol group there was no significant alteration except in the brain on the 21st day. The free fatty acids showed significant increases in the toddy treated animals on the 21st and 45th day. In the ethanol-administered rats, an increase in free fatty acids was seen only in the liver. The serum triglycerides did not show any significant change on administration of alcohol/toddy. The free fatty acids showed significant increases in treated groups on both days (21st and 45th) (Table 2).





Survival pattern of pups

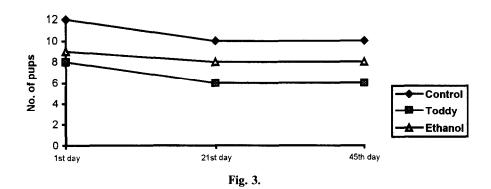


Table 1. Lipid profiles of various tissues and serum of offspring

	Liver		Kidney		Brain		Serum	
	21st	45th	21st	45th	21st	45th	21st	45th
Cholest	terol content m	$g 100 g^{-1}$ tissue	;					
CON	332 ± 14.8	294 ± 14.4	388 ± 15.4	376 ± 15.6	686 ± 18.8	875 ± 11.4^{b}	80 ± 1.61	83.1 ± 2.50
TOD	$420 \pm 12.3*$	305 ± 10.5^{ab}	377 ± 15.3	369 ± 18.8	$890 \pm 15.3*$	1011 ± 17.9* ^{ab}	81.9 ± 2.07	85.7 ± 1.16
ETH	$396 \pm 13.6\texttt{*}$	$390 \pm 14.3 \texttt{*}$	375 ± 15.6	352 ± 10.3	$866 \pm 16.7 \texttt{*}$	912 ± 19.1^{b}	85.5 ± 1.24	86.1 ± 1.70
Phosph	olipid content	mg 100 g ⁻¹ tiss	ue					
CON	2042 ± 118.7	2113 ± 112	1943 ± 103	1688 ± 118^{b}	1605 ± 114.6	2074 ± 114^{b}	90.8 ± 4.15	93.8 ± 5.26
TOD	$2406 \pm 113^{*a}$	2129 ± 114^{b}	2411 ± 113* ^a	1885 ± 117^{b}	$1715 \pm 112*$	2085 ± 118^{b}	$141.5 \pm 5.2^{*a}$	$138 \pm 5.50^{*a}$
ETH	2087 ± 116	2175 ± 119	2085 ± 121	1813 ± 112	1650 ± 119	2014 ± 116^b	$113\pm4.86\texttt{*}$	$114 \pm 5.21*$

*p < 0.05 between control and other groups.

 $^{a}p < 0.05$ between group II and group III.

 $b^{b}p < 0.05$ between 21st day and 45th day. Values expressed as mean \pm SE of six rats on 21st and 45th day.

The total bile acids showed a significant reduction in both the treated groups in 21st and a slight reduction in 45th day pups (Fig. 4). The activities of glucose-6phosphate dehydrogenase and malic enzyme were elevated significantly in the pups on the 21st day and elevated slightly on 45th day. Maximum activity was observed in toddy-exposed rats. (Fig. 5). The HMG Co-A Reductase activity was high in the toddy- and ethanol-treated groups. The ethanol group showed a slightly higher activity than the toddy group in 45th day pups (Fig. 6). Incorporation of labelled acetate into lipids showed an increased incorporation into cholesterol, phospholipids and triglycerides in 21st and 45th day pups, but there was a significant decrease in the incorporation to cholesterol ester content (Fig. 7).

DISCUSSION

Administration of toddy/ethanol decreased the weight gain of dams and this is reflected in the weight gain of

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	Liver		Kidney		Brain		Serum	
	21st	45th	21st	45th	21st	45th	21st	45th
Trigly	ceride content n	ng 100 g ⁻¹ tissue						
CON	217 ± 16.8	285 ± 12.6^{b}	231 ± 16.7	299 ± 21.5^{b}	614 ± 18.3	829 ± 18.6^{b}	12.1 ± 1.52	14.8 ± 2.16
TOD	$395 \pm 17.8^{*a}$	353 ± 12.9* ^a	$263 \pm 16.8*$	324 ± 18.9^{b}	$829 \pm 10.8^{*a}$	$864. \pm 18.5$	12.6 ± 1.28	16.4 ± 2.15
ETH	232 ± 17.9	299 ± 21.8^b	231 ± 15.2	319 ± 20.2^{b}	$680 \pm 17.8*$	837 ± 18.9^b	12.4 ± 2.10	18.5 ± 2.56
Free fa	atty acids mg 10	0 g ⁻¹ tissue						
	331 ± 14.6	421 ± 16.8^{b}	231 ± 28.3	246 ± 10.2	479 ± 14.52	525 ± 12.9^{b}	138 ± 15.1	143 ± 12.2
TOD	525±11.0*a	635±12.8* ^{ab}	288±16.3*a	357±13.6* ^{ab}	726±17.6*a	824 ± 12.5*ab	$232 \pm 18.2*$	$238 \pm 16.2*$
ETH	424 ± 10.8	$552 \pm 13.7^{*b}$	184 ± 25.6	254 ± 19.5^{b}	498 ± 18.6	543 ± 18.1^b	$239 \pm 20.1 *$	$248 \pm 18.9 \texttt{*}$

Table 2. Lipid profiles of various tissues and serum of offspring on 21st and 45th day

CON, Control; TOD, toddy; ETH, ethanol.

*p < 0.05 between control and other groups.

 $^{a}p < 0.05$ between group II and group III.

 $b^{b}p < 0.05$ between 21st day and 45th day. Values expressed as mean \pm SE of six rats.

	HDL-Ch	nolesterol	LDL + VLDL cholesterol		
	21st day	45th day	21st day	45th day	
Control	28.0 ± 1.04	31.4±2.92	52.0 ± 1.43	51.7±2.87	
Toddy	$20.9 \pm 1.63*$	$21.8 \pm 2.44*$	$60.9 \pm 2.86*$	64.0±2.94*	
Ethanol	$20.9 \pm 0.88*$	$16.8 \pm 1.11*$	$64.7 \pm 1.42*$	$69.3 \pm 1.47*$	

*p < 0.05 between control and other groups.

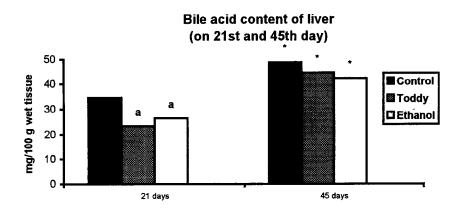


Fig. 4. a = p < 0.05 compared to control; * = p < 0.05 between 21 and 45 day pups.

pups and in their numbers. This observation is consistent with the earlier reports (Imai and Omoto, 1991; Breese *et al.*, 1993; Maldaner *et al.*, 1994). However, the survival rate of pups during the weanling period and after was not affected. This may be a result of the the low dose of alcohol administration.

In the present investigation, it is observed that ingestion of alcohol/toddy alters all the lipid parameters studied in the pups on the 21st day of lactation. These observations are in agreement with those of Druse (1981) and Lalitha *et al.* (1988), who have reported that cholesterol content of the fetus of alcoholic rats was elevated. Epidemiological studies on alcohol ingestion have shown a strong link between increased serum HDL and alcohol ingestion. Studies on the lipoprotein metabolism in prenatally alcohol-exposed pups are scarce. In this study we have observed low levels of serum HDL levels in 21 day and 45 day pups. This is an alarming factor since HDL is negatively correlated with coronary heart diseases (CHD). This may be a result of increased catabolism of HDL or its decreased biosynthesis. HDL is involved in the trasnsfer of cholesterol from tissues to the liver for degradation. Esterification of cholesterol by labelled studies was found to be decreased indicating lower HDL biosynthesis. Hence, lower HDL levels accords with the observed higher lipid levels in tissues.

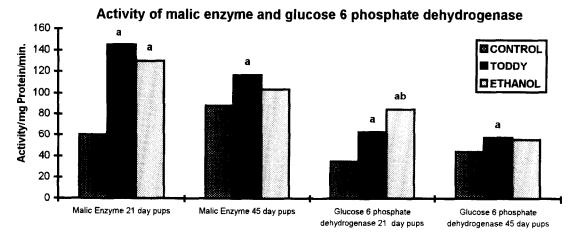
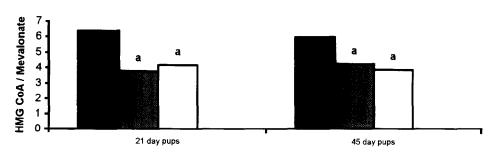
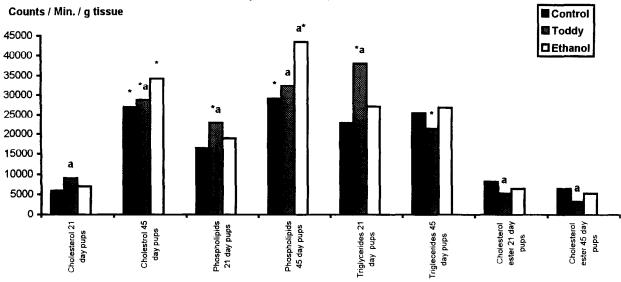


Fig. 5. a = p < 0.05 compared to control; b = p < 0.05 between treated groups.



Activity of HMG CoA reductase in the liver

Fig. 6. a = p < 0.05 compared to control.



14 C acetate incorporation in lipids (21 and 45 day pups)

Fig. 7. a = p < 0.05 compared to control; * = p < 0.05 between 21 and 45 day pups.

Only detailed studies on lipoprotein metabolism will reveal the underlying mechanism.

For 21 days after delivery, the pups were nursed by their own mothers. After that they were fed with a pelleted diet. It is known that milk from the alcoholic rat contains alcohol and its derivatives (Guerri and Sanchis, 1986). Therefore, the enhanced lipogenesis observed may be a result of the alcohol passed on to the pups through milk (Vilaro *et al.*, 1987). The elevation in the cholesterol content is a result of the enhanced lipogenesis, since there is an increase in the activities of lipogenic enzymes such as glucose-6-phosphate dehydrogenase and malic enzyme. The activity of the rate-limiting enzyme of cholesterol biosynthesis—HMG CoA reductase was also augmented. But the catabolism of cholesterol to bile acids was reduced, since hepatic bile acid content was depleted. This is in agreement with the report of Lakshmanan and Veech (1977) who have observed inhibition of cholesterol 7-alpha hydroxylase by ethanol.

On the 45th day of lactation there was a tendency for the cholesterol and phospholipid level to come down to the control level in the liver, but there were no significant changes in the activities of lipogenic enzymes or bile acid content. This reduction in cholesterol level was the result of a decrease in the synthesis of cholesterol since the activity of HMG Co A reductase was reduced.

In the brain and serum, even on the 45th day, that is when pups are no longer exposed to alcohol, hyperlipidemia is seen. It was shown previously that the peak period of lipid accumulation occurs in the rat brain in the first four weeks of postnatal life (Wells and Dittmer, 1967). So exposure to alcohol during gestation and lactation causes hyperlipidemia in the brain which cannot be reversed by abstaining from alcohol. This altered lipid pattern of the brain may affect the fluidity of membranes and neurotransmission (Lalitha *et al.*, 1988). This can partially explain the various abnormalities associated with FAS.

Analysis of the lipid profile reveals that toddy exerts more drastic changes than ethanol in the tissues. This is caused by increased synthesis and decreased catabolism of lipids in the toddy-exposed pups as evidenced by the activities of lipogenic enzymes, HMG CoA reductase, ¹⁴C acetate to lipids and decrease in the hepatic bile acids. The absolute alcohol intake of the toddy group and ethanol group was the same. So the non-alcoholic portion of the toddy is responsible for the augmented lipogenesis exerted by toddy in comparison with ethanol.

Toddy contains 2–5% alcohol (Child, 1972). Toddy contains 15% sucrose (Shamala and Sreekantiah, 1988). Hence, approximately the rats consumed $3.7 \,\mathrm{g\,kg^{-1}}$ body weight of sucrose day⁻¹. Sucrose is a hyperlipidemic agent (Naismith, 1971). Sucrose feeding was reported to produce higher levels of serum cholesterol when compared with glucose or starch. Reports from our laboratory have shown that sucrose-rich diet caused decreased degradation of cholesterol to bile acids in rats (Molly *et al.*, 1984). In addition to this, Ornoy and Cohen (1980) reported that a sucrose rich diet during pregnancy has teratogenic effects in rats. Jen *et al.* (1991) reported that sucrose reduces newborn weights.

In addition to sucrose, toddy contains B vitamins, ascorbic acid, minerals etc. (Shamala and Sreekantiah, 1988). Various studies have shown that ascorbic acid (Nambisan and Kurup, 1975; Saleena *et al.*, 1985) niacin (Thomas *et al.*, 1982), pyridoxin (Vijayammal and Kurup, 1978) are antiatherogenic vitamins whereas thiamine is atherogenic (Vijayammal and Kurup, 1981). So the concerted action of all the congeners may cause induced marked hyperlipidemia in toddy exposed pups in comparison with alcohol-exposed ones. It can therefore be concluded that, even though the alcohol content of toddy is very low, because of the presence of unfermented sugars and other congeners it is a potent hyperlipidemic agent. These studies throw light on the fact that pups born and nursed by alcoholic mothers have inborn hyperlipidemia, and it does not recover to normal levels even when the pups are not exposed to alcohol.

REFERENCES

- Abel, E. L. and Dintcheff, B. A. (1986) Effects of prenatal alcohol exposure on behaviour of aged rats. *Drug Alcohol Dependency* 16(4), 321–330.
- Bennet, C. J. and Franklin, N. L. (1967) Statistical Analysis in Chemistry and Chemical Industry, pp. 133. John Wiley and Sons Inc., New York.
- Breese, C. K., D'Costa, A., Ingram, R. L., Lenhan, J. and Soontag, W. E. (1993) Long term suppression of insulin like growth factor 1 in rats after in utero ethanol exposure: relationship to somatic growth. *Journal of Pharmacology Exposure Therapy* 264(1), 448–456.
- Charls, H. H. (1980) Alcoholism and malnutrition—Introduction to the symposium. *The American Journal of Clinical Nutrition* 33, 2705–2708.
- Child, R. (1972) *Toddy Products in Coconuts*. 2nd Edition, ed. R. Child, pp. 296–306. Longman Group Ltd, London.
- Clarren, S. K. and Smith, D. W. (1978) The fetal alcohol syndrome. New England Journal of Medicine 298, 1063-1067.
- Driscoll, C. D., Riley, E. P. and Meyer, L. S. (1985) Delayed taste aversion learning in preweanling rats exposed to alcohol prenatally. *Alcohol* 2(2), 277–280.
- Druse, M. J. (1981) Effect of maternal ethanol consumption in neurotransmitters and lipids in offspring. Neurobehav. Toxicology Terratol. 3, 81–87.
- Guerri, C. and Sanchis, R. (1986) Alcohol and acetaldehyde in rats milk following ethanol administration. *Life Science* **38**, 1543–1556.
- Imai, T. and Omoto, M. (1991) Effect of ethanol exposure beginning at an early stage on maternal rats and their offspring. Arukoru. Kenkyuto. Yakubutsulson 26(6), 544-568.
- Jen, K. L. C., Rochon, C., Zohong, S. and Whitcomb, L. (1991) Fructose and sucrose feeding during pregnancy and lactation in rats changes in maternal and pup fuel metabolism. *Journal of Nutrition* 121, 1999–2005.
- Jones, D. P. (1969) Effects of ethanol on lipid transport in man. In *Biochemical and Clinical Aspects of Alcohol Metabolism*, ed. V. M. Sardesai, pp. 86–94. Charles C. Thomas, Publisher, USA.
- King, J. C. and Fabro, S. (1983) Alcohol consumption and cigarette smoking: effects on pregnancy. *Clin. Obst. Gyne-cology* **26**(2), 437-448.
- Kornberg, A. and Horecker, B. L. (1960) Methods in Enzymology. First edition, eds S. P. Colowick and N. O. Kaplan, pp. 323. Academic Press, New York.
- Lakshmanan, M. R. and Veech, R. L. (1977) Short and long term effects of ethanol administration *in vivo* on rat liver HMG Co-A reductase and cholesterol 7-alpha hydroxylase activity. *Journal of Lipid Research* 18, 325–329.
- Lalitha, T., Kumar, K., Ramakrishnan, C. V. and Telang, S. D. (1988) Effect of maternal alcohol consumption on the lipid composition of CNS in the offspring. *Journal of Neu*rochemistry 50(5), 1346–1351.
- Lowry, O. H., Rosenbrough, N. J., Farr, A. L. and Randall, R. (1951) Protein measurement with Folin-Phenol reagent. *Journal of Biological Chemistry* 173, 265-275.

- Maldaner, F. H., Durgante, L. P., Murussi, M., Xavier, M. K., Dalmaz, C. and Ferrieira, M. B. (1994) Effect of chronic ethanol consumption on gestation and lactation in rats. *Integer. Physiology Behaviour Science* **29**(2), 141–150.
- Menon, P. V. G. and Kurup, P. A. (1976) Dietary fiber and cholesterol metabolism—effect of fiber rich polysaccharide from black gram (*Phaseolus mungo*) on cholesterol metabolism in rats fed normal and atherogenic diet. *Biomedicine* 24, 248-250.
- Molly, T., Leelamma, S. and Kurup, P. A. (1983) Effect of black gram fiber (*Phaseolus mungo*) on hepatic hydroxy methyl glutaryl Co A reductase activity, cholesterogenesis and cholesterol degradation in rats. *Journal of Nutrition* 113, 1104–1108.
- Molly, T., Leelamma, S. and Kurup, P. A. (1984) Effect of dietary carbohydrate on cholesterol lowering action of black gram fiber. *Journal of Biochemistry Biophysics* 21, 148– 150.
- Naismith, D. J. (1971) The Hyperlipidemic Property of Sucrose in 'Sugar', eds J. Yudkin, J. Edelman and L. Hough, Chapter 16, pp. 183-190.
- Nambisan, B. and Kurup, P. A. (1975) Ascorbic acid and glycosaminoglycan and lipid metabolism in guinea pigs fed normal and atherogenic diet. *Atherosclerosis* 22, 447-461.
- Norton, S. and Kotkoskie, L. A. (1991) Basic animal research. Recent Developments in Alcoholism 9, 95-115.
- Norton, S., Terranova, P., Na, J. Y. and Sancho-Tello, M. (1988) Early motor development and cerebral cortical morphology in rats exposed perinatally to alcohol. *Alcohol. Clinical Exposure Research* **12**(1), 130–136.
- Okishio, T., Nair, P. P. and Gordon, M. (1967) Studies on bile acids: The microquanitative separation of cellular bile acids by gas liquid chromatography. *Biochemistry Journal* 102, 654-658.
- Ornoy, A. and Cohen, A. M. (1980) Teratogenic effects of sucrose diet in diabetic and non diabetic rats. *Israel Journal* of Medical Science 16, 789–791.

- Rao, V. and Ramakrishnan, S. (1975) Indirect assessment of hydroxy methyl glutaryl CoA reductase activity in the liver tissue. *Journal of Clinical Chemistry* 21, 1523–1525.
- Saleena, M., Menon, P. V. G. and Kurup, P. A. (1985) Effect of administration of vitamin A, Ascorbic acid and nicotinamide adenine dinucleotide + flavin adenine dinucleotide on severity of myocardial infraction induced by isoproterenol in rats. *Indian Journal of Exp. Biology* 23, 500–504.
- Severo, O. (1960) In *Methods in Enzymology*. First edition, eds S. P. Colowick and N. O. Kaplan, pp. 739. Academic Press, New York.
- Shamala, T. R. and Sreekantiah, K. R. (1988) Microbiological and biochemical studies on traditional Indian palm wine fermentation. *Food Microbiology* 5, 157–162.
- Tanaka, H., Nasu, F. and Inomata, K. (1991) Fetal alcohol effects: decreased synaptic formations in the field CA3 of fetal hypocampus. *International Journal of Dev. Neu*roscience 9(5), 509-517.
- Thomas, V. E., Anila, M. and Kurup, P. A. (1982) Effect of nicotinic acid on concentration of aortic glycosaminoglycans in rats. *Indian Journal of Biochemistry Biophysics* 19, 150–151.
- Vijayammal, P. L. and Kurup, P. A. (1978) Pyridoxine and atherosclerosis: role of pyridoxine in the metabolism of lipids and glycosaminoglycans in rat fed normal and high fat, high cholesterol diets containing 16% casein. *Aust. Journal of Biological Science* **31**, 7-20.
- Vijayammal, P. L. and Kurup, P. A. (1981) Thiamine and metabolism of glycosaminoglycans in rats fed cholesterol free and cholesterol containing diet. *Indian Journal of Biochemistry Biophysics* 18, 134–138.
- Vilaro, S., Vinas, O., Remesar, X. and Herrera, E. (1987) Effects of chronic ethanol consumption on lactational performance in rat: mammary gland and milk composition and pups' growth and metabolism. *Pharmacology Biochemistry Behaviour* 27, 333-339.
- Wells, H. E. and Dittmer, J. C. (1967) A comprehensive study of postnatal changes in the concentration of the lipids of developing rat brain. *Biochemistry* 6, 3169-3175.